

Dehydrated Culture Media Bases / Media Supplements

# **Technical Information**

## Lysine Decarboxylase Broth

### Product Code: DM 1376

**Application:** Lysine Decarboxylase Broth is used for differentiating Salmonella Arizonae from the Bethesda Ballerup group of *Enterobacteriaceae*.

| ngredients                     | Gms / Litre |  |
|--------------------------------|-------------|--|
| Peptic digest of animal tissue | 5.000       |  |
| 'east extract                  | 3.000       |  |
| Dextrose                       | 1.000       |  |
| -Lysine hydrochloride          | 5.000       |  |
| Bromocresol purple             | 0.020       |  |
| inal pH ( at 25°C)             | 6.8±0.2     |  |

### **Principal & Interpretation**

Decarboxylase media were first described by Moeller <sup>(1-3)</sup> for detecting lysine and ornithine decarboxylase and arginine dihydrolase followed by Falkow who developed a lysine decarboxylase medium for the identification and differentiation of *Salmonella* and *Shigella* <sup>(4)</sup>. Lysine Decarboxylase Broth is used to study the decarboxylase reactions for members of *Enterobacteriaceae*. Lysine Decarboxylase Broth is also recommended by APHA <sup>(5, 6)</sup> and other workers <sup>(7, 8)</sup>. During the early stages of incubation, following inoculation, fermentation of dextrose by the organisms leads to acid production, which causes a colour change of the bromocresol purple indicator to yellow. The acidic condition thus generated stimulates decarboxylase activity, which leads to decarboxylation of lysine to cadavarine. The alkaline conditions generated due to cadaverine production cause the bromocresol purple indicator (changed to yellow) to revert back to purple colour. If the organisms do not produce decarboxylase enzyme, the colour of the medium remains yellow. Dextrose non-utilizers will not show any change in the medium colour. Use light inocula and do not read the tests after 24 hours incubation, as some organisms require longer incubation time of upto 4 days.

### Methodology

Suspend 14.02 grams of powder media in 1000 ml distilled water. Shake well & heat, if necessary to dissolve the medium completely.

Dispense 5 ml amount into screw-capped test tubes. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes.

# **Quality Control**

#### Physical Appearance

Light yellow to greenish yellow homogeneous free flowing powder

Colour and Clarity of prepared medium Purple coloured clear solution without any precipitate

#### Reaction

Reaction of 1.4% w/v aqueous solution at 25°C. pH : 6.8±0.2

pH range: 6.6-7.0

#### Cultural Response/Characteristics

DM 1376: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.(Inoculated tubes are overlayed with sterile mineral oil).





| Organism                          | Inoculum (CFU) | Lysine decarboxylation           |
|-----------------------------------|----------------|----------------------------------|
| Citrobacter freundii ATCC 8090    | 50-100         | Variable reaction                |
| Escherichia coli ATCC 25922       | 50-100         | Variable reaction                |
| Enterobacter aerogenes ATCC 13048 | 50-100         | Positive reaction, purple colour |
| Klebsiella pneumoniae ATCC 13883  | 50-100         | Positive reaction, purple colour |
| Proteus mirabilis ATCC 25933      | 50-100         | Negative reaction, yellow colour |
| Proteus vulgaris ATCC 13315       | 50-100         | Negative reaction, yellow colour |
| Salmonella Arizonae ATCC 13314    | 50-100         | Positive reaction, purple colour |
| Salmonella Paratyphi A ATCC 9150  | 50-100         | Negative reaction, yellow colour |
| Salmonella Typhi ATCC 6539        | 50-100         | Positive reaction, purple colour |
| Serratia marcescens ATCC 8100     | 50-100         | Positive reaction, purple colour |
| Shigella dysenteriae ATCC 13313   | 50-100         | Negative reaction, yellow colour |

### Storage and Shelf Life

**Dried Media:** Store below 30°C in tightly closed container and use before expiry date as mentioned on the label. **Prepared Media:** 2-8<sup>0</sup> in sealable plastic bags for 2-5 days.

### Further Reading

1. Moeller V., 1954, Acta. Pathol. Microbiol. Scand., 34:102.

2. Moeller V., 1954, Acta. Pathol. Microbiol. Scand., 34:259.

3. Moeller V., 1955, Acta. Pathol. Microbiol. Scand., 36:158.

4. Falkow, 1958, Am. J. Clin. Pathol., 29:598.

5. Eaton A. D., Clesceri L. S., Rice E. W. and Greenberg A. W., (Eds.), 2005, Standard Methods for the Examination of Water and Wastewater, 21st Ed., APHA, Washington, D.C.

6. Downes F. P. and Ito K., (Eds.), 2001, Compendium of Methods for the Microbiological Examination of Foods, 4th Ed., American Public Health Association, Washington, D.C.

7. Isenberg (Ed.), 1992, Clinical Microbiology Procedures Handbook, Vol. 1. American Society for Microbiology, Washington, D.C.

8. FDA Bacteriological Analytical Manual, 2005, 18th Ed., AOAC, Washington, DC.

### **Disclaimer**:

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